# A SYSTEMS APPROACH TO CELLULAR SIGNAL TRANSDUCTION

Jeremy E. Purvis

### A DISSERTATION

 $_{\mathrm{in}}$ 

### Genomics and Computational Biology

Presented to the Faculties of the University of Pennsylvania in Partial Fulfillment of the Requirements for the Degree of Doctor of Philosophy



Platelets adhering to collagen, triggering intracellular calcium release

Sean Maloney | *unpublished data* 

Reactions governing the DNA damage response give rise to pulses of activated p53



Reactions governing coagulation give rise to waves of calcium in a developing thrombus



Lahav et al., 2004. Furie and Furie, 2008.

A single AA substitution in the epidermal growth factor receptor leads to constitutive growth signals



Expression of c-myc and ras<sup>D</sup> in transgenic mice dramatically accelerates tumor growth



Choi et al., 2007. Sin et al., 1987.

Computational approaches elucidate two clinically-relevant signaling responses

Oncogenic signaling through the epidermal growth factor receptor Ravi Radhakrisnan (PI) Yingting Liu

Andrew Shih Neeraj Agrawal

 Calcium and phosphoinositide signaling in human platelets
Scott Diamond (PI)
Skip Brass (PI)
Manash Chatterjee





### New modeling methods afford specific design goals

Steady-state kinetic modeling constrains cellular resting states Ravi Radhakrisnan (PI) Scott Diamond (PI)



 Dynamic neural network modeling predicts responses to multi-input signals
Scott Diamond (PI) Manash Chatterjee





Phosphotyrosine
Signaling in the
Epidermal Growth
Factor Receptor

Liu et al. (2007). *Ann Biomed Eng* 35:1012-25 Purvis et al. (2008). *Biotechnol Prog* 24:540-53 Shih et al. (2008). *Mol Biosyst* 4:1151-9.

- Addresses the question of why certain cancer cell lines are responsive to tyrosine kinase inhibitor (TKI) therapy
- Seeks to resolve how structural alterations in the receptor are communicated as downstream growth or survival signals

Differences in receptor signaling determine cell fate

Growth factor signaling influences cell decisions through receptor activation and trafficking, effector processing, and transcription

Patterns of tyrosine phosphorylation encode varying signals mediated through adaptor proteins



JAK2

Grb2 STAT3/5

GAB-1

PTP1

→ Akt

**ERK1/2** 

pY1068

pY1086

pY1148

pY1173

Mutant behavior resolved through molecular dynamics and molecular docking

Molecular dynamics simulations showed that the drug sensitizing mutation (L858R) of EGFR stabilizes the active conformation

Andrew Shih

Docking simulations showed that L858R had an increased affinity for the drug and a preference for certain phosphorylated tyrosine residues Yingting Liu





### Increased flux through Y1068 causes a relative gain in Akt activation



Table 1.	Parametric	Differences	between	WT	and	Mutant	EGFR
Systems I	Inferred from	m Experime	ents				

parameter	WT	L834R	del	reference
$K_{\rm M}^{\rm ATP}$	$5.0\mu\mathrm{M}$	10.9 μM	$129 \mu M$	3
$K_{\rm I}^{\rm erlotinib}$	17.5  nM	6.25 nM	$3.3 \ \mu M$	3
$K_{M}^{Y1068}$	$265 \mu M$	13.3 μM	$130 \mu M$	14, 57, 58
$K_{M}^{\tilde{Y}_{1173}}$	$236\mu\mathrm{M}$	$200  \mu M$	$300 \mu M$	14, 57, 58
$k_{cat}^{\dot{Y}\hat{1}068}$	$0.29 \text{ s}^{-1}$	$0.24 \text{ s}^{-1}$	$0.2 \ s^{-1}$	14, 54, 57, 58, 59
$k_{cat}^{Y1173}$	$0.25 \ s^{-1}$	$0.21 \text{ s}^{-1}$	$0.22 \ s^{-1}$	14, 54, 57, 58, 59



Purvis, Ilango, and Radhakrishnan (2008). Sordella et al. (2004)



Inhibitor



Inhibitor

Multiscale modeling offers mechanistic explanation for oncogene addiction

Response of L858R to TKI therapy is an example of "oncogene addiction"

Sensitivity may be used to identify probable resistance mechanisms

Method provides a link between mutational status and kinetic properties of the system

Akt, and not Erk, is the major mediator of cell survival in oncogenic mutants of EGFR







# II. Calcium Signaling in Human Platelets

Purvis et al. (2008). *Blood* 112:4069-4079 Purvis et al. (2009). *PLoS Comput Biol* Chatterjee et al. (2009). *submitted* 

- Platelet activation is central to the 1.75 million heart attacks and strokes that occur annually in the US
- Quantitative models can make accurate predictions of patient-specific risks and biological mechanism

# Platelet signaling regulates hemostasis and thrombosis

1



### Normal circulation





<sup>1</sup> Brubaker (2006). <sup>2</sup> Chatterjee, Purvis, and Diamond (2009)



### Opposing kinetic processes balance calcium in the platelet



Cytosolic calcium concentration ( $[Ca^{2+}]_{cyt}$ )  $\approx$  100 nM Lumenal calcium concentration ( $[Ca^{2+}]_{dts}$ )  $\approx$  200-650  $\mu$ M Extracellular calcium concentration ( $[Ca^{2+}]_{dts}$ )  $\approx$  1 mM Lumenal fraction appears to occupy ~0.1 to 10% of reactive volume

SERCA3 Calcium ATPase

### IP<sub>3</sub> Receptor Channel



Ebbeling et al., 1992. *Blood* 80:718. Dode *et al*, 2002. Sneyd and Dufour, 2002.

### Fix topology and sample Ca<sup>2+</sup> space



### Preferred concentrations/structures



Purvis, Chatterjee, Brass, and Diamond, 2008



<sup>1</sup> Purvis, Chatterjee, Brass, and Diamond, 2008. <sup>2</sup> Wilson, Neufeld, and Majerus, 1985.

Modulation of PKC activity by second messengers, Ca<sup>2+</sup> and DAG Activated PKC translocates to PM and phosphorylates PLC-β

Negative feedback necessary for synchronized responses







# Resting modules are merged to form a full kinetic model



Purvis, Chatterjee, Brass, and Diamond, 2008. Blood 112:4069.





### Dose response to ADP



### **Population Responses:**

Smooth, average responses are observed when cells are pooled

Purvis, Chatterjee, Brass, and Diamond, 2008.

### Individual platelets exhibit noisy calcium release behavior













### Small cell volume gives rise to stochastic response



Ca<sup>2+</sup> peak intervals for observed

# Platelet calcium signaling is stochastic

Platelet volume: ~ 8 fL

1 nM = 4 molecules per platelet

Ca<sup>2+</sup> response as a function of cell volume



# **Platelet Volume**

Purvis, Chatterjee, Brass, and Diamond, 2008.

Platelet model is extensible to comprehensive models of coagulation

 Quantitatively understand Ca<sup>2+</sup> balance/release mechanisms

 Recapitulate platelet response to multiple agonists

 Integrate into larger scale models of coagulation

 Predict blood function and pharmacological sensitivity











III. Steady-State Kinetic Modeling Constrains Cellular Resting States

> Purvis et al. (2008). *Blood* 112:4069-4079 Purvis et al. (2009). *PLoS Comput Biol*

- Signaling systems are robust to small perturbations and have a steady state
- Large kinetic models have many unknown values that are possibly correlated



- Often, topology is know but concentrations are not
- Each molecule occupies a separate linear dimension
- Each module comprises a subspace of the full concentration space
- Modules that share a species have intersecting subspaces

Purvis, Radhakrishnan, and Diamond (2009). PLoS Comput Biol 5:e1000298

### Homeostasis constraint and PCA reduce allowable cellular configurations



Purvis, Radhakrishnan, and Diamond (2009). PLoS Comput Biol 5:e1000298

### Resting modules are merged to find global time-dependent solution





### Alternate steady state profiles reveal `rigid´ and ` flexible´ concentrations



	`Rigid' nodes:	`Flexible' nodes:
Values fixed during estimation	$\begin{bmatrix} [Ca^{2+}]_i \\ PIP_2 \\ GTP \\ PIC-B \end{bmatrix}$	PKC G <sub>q</sub> -GTP P2Y <sub>1</sub> SFRCA

Purvis, Chatterjee, Brass, and Diamond, 2009. Blood 112:4069

# A realistic description of signaling requires a multitude of reactions





IV. Dynamic neural network modeling predicts responses to multi-input signals

Chatterjee, Purvis, and Diamond (2009) *submitted* 

- Cells must respond to multiple potent signals *in vivo*
- Addresses question of how multiple simultaneous signals are integrated
- Sensitive enough to resolve differences in signaling among individuals

# Multiple signaling pathways converge on intracellular calcium release



### Signal inputs are normalized according to EC<sub>50</sub>



### 4 agonists states:

 $0 \times EC_{50}$  $0.1 \times EC_{50}$  $1 \times EC_{50}$  $10 \times EC_{50}$ 

Pladelet agonist dose responses

Selecting low, moderate, and high doses for each agonist **normalizes** the input signals and **captures the dynamic range** of each agonist

### A high-throughput assay measures platelet response to combinatorial inputs



NN is trained on responses to all pairwise combinations of agonist

Network trained on all pairwise combinations of 6 agonists at 3 concentrations

(6 choose 2)  $\times$  3<sup>2</sup> + 19 = 154 combinations



Define synergy as the difference between the integrated calcium transient for the combined response and the integrated area for the individual responses







# NN predicts entire 6-dimensional platelet response

- NN made 4077 predictions of synergy among all combinations of 6 agonists at 3 doses
- 45 dissimilar conditions were chosen for experimental verification



### Sample (n = 45) of global response space identifies high-synergy conditions



# NN predicts sequential responses and downregulation



ExperimentSimulation

### NN models provide a compact, patient-specific predictive tool



Train NN to match calcium profile for specific donor:

### Weight values for donor-trained NN:



# Acknowledgements

Advisors Scott Diamond Ravi Radhakrishnan

Platelet and Modeling Expertise Skip Brass Talid Sinno

Thesis Committee Chair Lyle Ungar

Genomics and Computational Biology Graduate Group Maya Bucan Warren Ewens Junhyong Kim Shane Jensen Jeff Saven

### Blood Systems Biology Group

Sean Maloney Manash Chatterjee Matthew Flamm Dan Jaeger

Software & Resources

libSBML SBToolbox<sup>2</sup> SBML Shorthand BioNetGen<sup>™</sup> DOQCS BRENDA

Funding R01-HL-56 621 R33-HL-87 317 T32-HG000046



