Check for updates

OPEN ACCESS

EDITED BY Zane Lybrand, Texas Woman's University, United States

REVIEWED BY Mathew V. Jones, University of Wisconsin-Madison, United States Kioko Rubi Guzman-Ramos, Autonomous Metropolitan University, Lerma, Mexico

*CORRESPONDENCE David F. Meaney dmeaney@seas.upenn.edu

SPECIALTY SECTION This article was submitted to Cellular Neuropathology, a section of the journal Frontiers in Cellular Neuroscience

RECEIVED 24 June 2022 ACCEPTED 25 October 2022 PUBLISHED 24 November 2022

CITATION

Schumm SN, Gabrieli D and Meaney DF (2022) Plasticity impairment alters community structure but permits successful pattern separation in a hippocampal network model. *Front. Cell. Neurosci.* 16:977769. doi: 10.3389/fncel.2022.977769

COPYRIGHT

© 2022 Schumm, Gabrieli and Meaney. This is an open-access article distributed under the terms of the Creative Commons Attribution License

(CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Plasticity impairment alters community structure but permits successful pattern separation in a hippocampal network model

Samantha N. Schumm¹, David Gabrieli¹ and David F. Meaney^{1,2*}

¹Department of Bioengineering, School of Engineering and Applied Sciences, University of Pennsylvania, Philadelphia, PA, United States, ²Department of Neurosurgery, Penn Center for Brain Injury and Repair, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA, United States

Patients who suffer from traumatic brain injury (TBI) often complain of learning and memory problems. Their symptoms are principally mediated by the hippocampus and the ability to adapt to stimulus, also known as neural plasticity. Therefore, one plausible injury mechanism is plasticity impairment, which currently lacks comprehensive investigation across TBI research. For these studies, we used a computational network model of the hippocampus that includes the dentate gyrus, CA3, and CA1 with neuronscale resolution. We simulated mild injury through weakened spike-timingdependent plasticity (STDP), which modulates synaptic weights according to causal spike timing. In preliminary work, we found functional deficits consisting of decreased firing rate and broadband power in areas CA3 and CA1 after STDP impairment. To address structural changes with these studies, we applied modularity analysis to evaluate how STDP impairment modifies community structure in the hippocampal network. We also studied the emergent function of network-based learning and found that impaired networks could acquire conditioned responses after training, but the magnitude of the response was significantly lower. Furthermore, we examined pattern separation, a prerequisite of learning, by entraining two overlapping patterns. Contrary to our initial hypothesis, impaired networks did not exhibit deficits in pattern separation with either population- or rate-based coding. Collectively, these results demonstrate how a mechanism of injury that operates at the synapse regulates circuit function.

KEYWORDS

traumatic brain injury, cellular networks, memory impairment, plasticity, patterned structure

Introduction

Traumatic brain injury (TBI) is a debilitating condition that involves dysfunction across diverse neural circuitry. Often a result of impacts to the head, TBI is pervasive with up to 2.5 million cases recorded in 2014 (Cancelliere et al., 2017; Taylor et al., 2017). The incidence of TBI has risen along with increasing societal awareness of the issue (Prince and Bruhns, 2017), owing in part to the effects of concussion on adolescents and young adults (Giza et al., 2017). Fortunately, most patients with mild TBI recover relatively quickly (<3 months) (Ruff, 2005); however, others experience prolonged symptoms, including headaches, reduced processing speed, and attention or memory impairments (Ruff, 2005; Prince and Bruhns, 2017). Recent evidence points to the myriad consequences of TBI across the spectrum, ranging from poor outcome in severely brain injured patients (Kondziella et al., 2022), the broad and persisting impairments that can occur after mild TBI (Lopez et al., 2022), how TBI continues to increase risk for other neurological disorders which include epilepsy (Mariajoseph et al., 2022) and the substantial employment and economic consequences of TBI across the severity spectrum (Gaudette et al., 2022).

Memory deficits are among the most common and potentially detrimental complaints among TBI patients (Mcallister et al., 2001; Nicholl and Curt LaFrance, 2009; McAllister, 2011). The hippocampus is a well-studied brain structure known especially for its contributions to memory processes (Sanders et al., 2015; Jeffery, 2018; Genon et al., 2021; Slotnick, 2022). Earlier work has shown that the hippocampus is vulnerable to TBI and easily damaged (Kotapka et al., 1994; Raghupathi, 2004; McAllister, 2011; Paterno et al., 2017). Behavioral studies in rodents have proved hippocampal involvement in both working and episodic memory and that deficits occur after TBI across the severity spectrum (Smith et al., 2015; Paterno et al., 2017). Spatial memory, a subtype of episodic memory, has also been extensively studied with in vivo TBI models, which exhibit protracted dysfunction after mild injury (Dawish et al., 2012; Paterno et al., 2018).

The prevailing theory of memory describes three distinct phases—encoding, maintenance, and retrieval (Tulving, 1985; Josselyn et al., 2015). Encoding is the construction of a persistent neural representation, or memory, of an experience, maintenance entails preservation of the memory over time, and retrieval is the active process of recall or accessing the memory anew. The hippocampus is involved in all three procedures (Paterno et al., 2017), but precisely how TBI perturbs these three phases remains unclear. The process of forming memories is supported by synaptic plasticity, a mechanism by which circuits are strengthened or weakened. In classical electrophysiology, such enduring, use-dependent increases in synaptic strength are encompassed by the phenomenon of long-term potentiation (LTP), or the enhancement of synaptic transmission efficiency. After TBI, several groups have demonstrated changes in synaptic plasticity and failures in LTP (Albensi et al., 2000; Sanders et al., 2000; Schwarzbach et al., 2006; Aungst et al., 2014; Wilson et al., 2016; White et al., 2017), especially in area CA1 of the hippocampus (Schwarzbach et al., 2006; Cohen et al., 2007), suggesting plasticity impairment may underlie post-injury behavioral deficits in memory tasks. The inability to induce LTP is associated with reduced CamKII phosphorylation and synaptic protein disruption, which together represent a lower capacity for synaptic potentiation (Schwarzbach et al., 2006; Vogel et al., 2017). If LTP impairment represents a potentiation deficit and potentiation undergirds memory formation, we would anticipate encoding problems to ensue after injury. Surprisingly, there is no consensus about which phase of the memory process is most disrupted after injury.

Beyond the biological basis of memory and the disruption posed by TBI, the adaptation of microcircuit architecture through learning remains largely unaddressed in the existing literature. One tool used at the macroscale is modularity for community detection in large networks. Communities are clusters of nodes with connections to one another that facilitate performing a collaborative function (Sporns and Betzel, 2016). The division of the brain into functional subnetworks is well-supported at the macroscale (Sporns and Betzel, 2016). Specific to learning, one group examined how networks evolve over the course of learning through dynamic community realignment (Bassett et al., 2011). How the concepts of modularity and learning integrate in microscale circuits is unknown. While attention and learning are often studied in macroscale brain networks, there are few existing studies of learning in biologically derived microscale neural networks (Izhikevich, 2006; Guise et al., 2015; Chavlis et al., 2017; Gabrieli et al., 2020a). A few groups have documented how connectivity adapts with stimulation and development in vitro (Whittington and Traub, 2003; Draguhn and Buzsáki, 2004; Penn et al., 2016), and some models have considered learning-related inputoutput relationships. However, these are limited by either a lack of plasticity or specific physiological network structure. For instance, Chavlis and colleagues analyzed the effect of dendritic atrophy on pattern separation in a computational model of the dentate gyrus (Chavlis et al., 2017); however, since the model does not incorporate plasticity, the results do not invoke classical potentiated learning. Examining community structure in a computational model of the hippocampus facilitates finer resolution analysis than could otherwise be obtained experimentally because we can observe the evolution of thousands of neurons over time. Furthermore, we can study the effects of an isolated mechanism of injury that has circuity-level implications. Among many possible outcomes of secondary injury sequelae, plasticity impairment can be directly linked to learning and memory dysfunction. Reports of learning-dependent network changes in this important, memory-related microcircuit are currently lacking.

In these studies, we use a model of three integrated subregions of the hippocampal formation (namely, the DG, CA3, and CA1) that comprise the classical trisynaptic circuit. The model was constructed according to known electrophysiology and anatomical connectivity data. Although TBI likely includes a plethora of mechanisms, we isolated the effects of impaired potentiation. We simulate one effect of mild TBI as STDP impairment by reducing potentiation in the circuit and establish that this deficit reduced activity and broadband power in the network (Schumm et al., 2022). Here we extend those results by demonstrating how STDP impairment affects the structural network, focusing specifically on community organization. STDP impairment causes realignment among excitatory neurons in CA3. We also implement a learning paradigm using overlapping input patterns to study pattern separation across the hippocampal subregions. Networks with STDP impairment exhibit minor learning impairments but no pattern separation deficits, despite significant activity differences and modified community structure. These results should be interpreted given the limitations of this work; the computational network model is not full-scale, and the mechanism of injury (plasticity impairment) does not encompass the full scope of potential dysfunction post-TBI. However, the approach provides an opportunity to isolate the effects of potentiation deficits in a network model of all three primary sub-regions of the hippocampus.

Materials and methods

Briefly, the model focuses on the dentate gyrus (DG), CA3, and CA1 as the primary subregions of the hippocampal formation (**Figures 1A**,C). The areas follow a primarily feedforward topology with the DG sending projections to CA3 which terminates in CA1 (a summary of the modeling methods follows. See Schumm et al., 2022 for more information about the model parameters).

Network structure and model dynamics

The network is a system of nodes that represent neurons and edges that designate the connections between them. For each point neuron, we applied the Izhikevich integrate-and-fire neuron model, which uses the following system of differential equations to determine the spiking behavior of a neuron over time (Izhikevich and Edelman, 2008):

$$Cv' = k(v - v_r)(v - v_t) - u + I$$
(1)

$$u' = a \left[b \left(v - v_r \right) - u \right] \tag{2}$$

if
$$v \ge v_p$$
, then
$$\begin{cases} v = c \\ u = u + d \end{cases}$$
 (3)

Where v is the membrane potential in millivolts (mv), and u is the recovery variable. C is the membrane capacitance (pF), v_r is the resting membrane potential, v_t is the threshold potential, and v_p is the membrane potential at the peak of the spike. Iis current in picoamperes (pA). The dimensionless parameters a, b, c, d, and k are adjusted to represent different subtypes of neurons. The current (I) aggregates receptor-based ionic currents, including AMPA, NMDA, and GABA-A receptors, and 1 Hz noise input that drives the network and follows a gamma distribution (k = 2, $\theta = \frac{1}{2}$) (Izhikevich and Edelman, 2008; Gabrieli et al., 2020b; Schumm et al., 2020, 2022).

There are 10 different types of neurons represented in the model across the three anatomical subregions (Schumm et al., 2022). The dentate consists of granule cells, mossy cells, basket cells, and interneurons. Areas CA3 and CA1 each have pyramidal cells, basket cells, and interneurons with parameters specific to that subregion. Inhibitory neurons account for approximately 10% of the neurons in each subnetwork (Woodson et al., 1989; Aika et al., 1994; Bezaire and Soltesz, 2013; Bezaire et al., 2016). The subtypes have characteristic electrophysiology and connectivity, which are represented through functional and structural features of the model, respectively. Broadly, the connectivity of the hippocampus follows a feedforward architecture (see Schumm et al., 2022 for detailed connectivity information). Granule cells, the principal excitatory neurons of the dentate, synapse onto CA3 neurons but have no connections to one another under physiological conditions. CA3 pyramidal cells are known to have a relatively high proportion of recurrent collaterals, but the majority of their axons project to CA1 pyramidal cells. In total, there are 8,885 neurons in the model, which converts to a scale of approximately 1:185 principal neurons in the rat hippocampus.

Plasticity implementation and impairment

The model incorporates two primary forms of synaptic plasticity—spike-timing-dependent plasticity (STDP) and homeostatic plasticity (HSP). STDP is a form of order-dependent Hebbian learning. The process relies on precise spike timing between neurons and strengthens synapses when neurons fire causally (i.e., when the upstream neuron fires before the downstream neuron) (Feldman, 2012). Synaptic strengthening and weakening occur according to the following equation (Effenberger et al., 2015):

$$\Delta w(w) = \begin{cases} A_{+}(w) \exp\left(-\frac{t_{post}-t_{pre}}{\tau}\right) & \text{if } t_{post}-t_{pre} > 0\\ A_{-}(w) \exp\left(-\frac{t_{post}-t_{pre}}{\tau}\right) & \text{if } t_{post}-t_{pre} \le 0 \end{cases}$$
(4)



Where w is the weight of the connection between two neurons. A_+ and A_- determine the magnitude of maximal synaptic change. The A_+/A_- ratio is often biased toward strengthening and equaled 1.05 in this work (Song et al., 2000). τ is the plasticity time constant and was approximated as 20 ms (Song et al., 2000). Finally, t_{pre} and t_{post} are the timing of preand post-synaptic spikes, respectively. Similar to previous models, plasticity applied to excitatoryto-excitatory synapses only (Song et al., 2000). While there are documented cases of inhibitory plasticity, inhibitory STDP is highly variable (Lu et al., 2007; Caporale and Dan, 2008), making it difficult to implement in the model without further empirical study within this circuit. To stabilize connection weights in the network (Zenke et al., 2017), we incorporated synaptic scaling, a specific form of HSP that operates at the level of individual neurons (Tetzlaff et al., 2011). The activity of each neuron is compared to a target firing rate, and all the synapses of the neuron are modified to shift the actual firing rate closer to the target firing rate (Turrigiano et al., 1998; Turrigiano and Nelson, 2004). The following equation describes a threshold formulation of HSP adapted from (Effenberger et al., 2015):

$$if |(v_o - v_t)/v_t| > 0.50$$
(5)

$$\Delta w(w) = -\frac{\gamma}{W_{max}} \left(\frac{v_o - v_t}{v_t}\right)(w^2) \tag{6}$$

Where *w* is the weight of connection, γ is the dimensionless rate of change and equals 10^{-8} in these studies, v_o is the observed firing rate, v_t is the target firing rate, and W_{max} is the maximum excitatory weight of that neuron subtype. The function has a threshold such that synaptic weights are adjusted for neurons with firing rate change greater than 50% of their target firing rate (v_t) over the course of 120 s. This threshold ensures that the network continues to adapt with STDP without creating neurons with unconstrained, runaway activity.

STDP is associated with the well-studied phenomenon of long-term potentiation (LTP) observed in brain slice electrophysiology (Feldman, 2012). LTP describes the prolonged increase in synaptic efficacy of a circuit and is believed to support learning at the organismal level. TBI leads to deficits in spatial learning and LTP (Albensi et al., 2000; Aungst et al., 2014; Paterno et al., 2017; White et al., 2017), especially within CA1 of the hippocampus (Schwarzbach et al., 2006; Cohen et al., 2007). We sought to mimic a plasticity deficit and effects of mild TBI by altering the STDP algorithm in our model. To achieve this impairment, we reduced the maximal amount of potentiation in the model by 10% (A+ = 0.9 instead of 1.0 in Equation 4 above) (Figure 1B). In our previous work, we demonstrated that this modest decrement contributed to significant decreases in firing rate and signal power in impaired networks (Figures 1D,E). Simulations ran for 20 min without HSP to expedite synaptic settling and then 30 min with HSP. Simulations with STDP impairment were run for an additional 30 min. Analysis was performed on the final 5 min of simulation time for both baseline and impaired networks.

Modularity analysis for community detection

Large network architectures can be partitioned into several subnetworks that perform specialized functions (**Figures 2A,B**). These modules or communities generally contain densely connected nodes that are more weakly connected to other nodes outside the module. There are many methodological options for conducting community detection in networks (Sporns and Betzel, 2016). Since our networks are directed, weighted, and

signed in addition to being large (more than 3,000 nodes), we required algorithms that could accommodate networks with this combination of characteristics. Modularity is one common technique used to detect the community structure of a network. Reorganizing the original matrix based on its underlying community structure takes several steps that we implemented with functions from the publicly available Brain Connectivity Toolbox (Rubinov and Sporns, 2010). Overall, we followed a procedure of modularity maximization which seeks to find the optimal network partition that maximizes the modularity quality function (Q) (Sporns and Betzel, 2016):

$$Q(\gamma) = \frac{1}{2m} \sum_{i,j} \left[a_{ij} - \gamma p_{ij} \right] \delta(\sigma_i, \sigma_j)$$
(7)

Where $a_{i,j}$ is the number of connection between modules i and j, $p_{i,j}$ is the expected number of connections between modules i and j according to a null model, 2m is the total number of connections, γ is the resolution parameter, and $\delta(\sigma_i, \sigma_j)$ is the Kronecker delta function.

The resolution parameter (γ) determines the scale of the modules that can be detected such that larger modules are detected with smaller gamma values. For hypothesis testing, a null model was generated by rewiring the original matrix while preserving the original input and output degree distributions. In gamma optimization, modularity (Q) is calculated for both experimental and null matrices across a sweep of gamma values (Figure 2C). The value of gamma that yields the largest difference in Q between the experimental and null matrices was used for subsequent steps. Gamma was optimized for minute 26 of each baseline simulation and held constant for ensuing timepoints and impaired models. With the optimized gamma parameter, we partitioned the matrix into communities many times to ensure robustness (Blondel et al., 2008). An association matrix was generated from the partition ensemble to obtain the consensus community partition. A null association matrix was also generated from a permuted partition ensemble, which is generated by permuting each column of the original partition ensemble. This null association matrix was used to threshold the experimental association matrix, thereby removing low weight connections. Consensus clustering produces an optimal partition with community assignments for each node, or neuron. Based on these assignments, the original matrix was reordered to represent the underlying community structure. We report the modularity (Q), the number and size of modules, and the composition of modules in the hippocampal networks.

Learning and pattern separation

The hippocampus plays a key role in the broad functions of learning and memory, which depend on long-lasting, if not permanent, changes to network circuitry. These network modifications are supported by plasticity mechanisms like STDP



that encode persistent responses to network stimulation. More specifically within the hippocampal formation, the dentate is known to execute the function of pattern separation, a crucial learning task in which similar incoming patterns become increasingly different from one another as they exit the network. In contrast, area CA3 with its recurrent collateral structure better supports pattern completion whereby partial pattern representations are completed as they pass through the network.

Although there are many ways to test learning in a neural network, given the size of our networks (>8,000 nodes), an unsupervised learning algorithm was preferable to a supervised approach, so we evaluated learning with a similar method to our previous work (Gabrieli et al., 2020a). To summarize this method, we applied two protocols to assess learning. During

training, the network was stimulated and able to adapt with plasticity to encode responses to periodic input over 30 min. During *testing*, static networks were stimulated for 6 min. Networks were tested before and after training to determine how training modified the network response.

The networks were first settled as described previously for 30 min of simulation time with 1 Hz noise and then trained with exogenous 1 Hz stimulus. For each of two patterns, we simultaneously stimulated a set of 200 input neurons in the DG and measured the response in all three subregions. The input patterns overlapped by 50% with 100 neurons that were common to both patterns and 100 neurons that were unique to each stimulus. The simulation ran for an additional 30 min with 1 Hz noise and 1 Hz stimulation of each pattern to encode the

activity response before the network was tested. The response was measured in the 200-ms epoch immediately following stimulation of either pattern 1 or pattern 2. Since learning is defined by training-dependent changes in network activity, we tested the response of untrained and trained networks to the two input patterns in order to determine which neurons augmented their activity after the training period. The activity of each neuron post-training was normalized by its activity before training to account for neurons with inherently high activity. The 200 neurons that increased their firing the most from untrained levels comprised the desired, target response. The remaining neurons made up the off-target response where increases in activity are undesirable. Thus, the response for each subregion consists of a target component of 200 neurons that respond maximally to the stimulus and an off-target component of the remaining principal neurons. The signal-to-noise ratio was measured as the ratio of the target to off-target response. Finally, this paradigm was repeated for two training conditions. One set of networks was trained under baseline conditions, and another set of networks underwent training with STDP impairment to test whether reduced potentiation interferes with the ability to encode patterned responses.

To evaluate pattern separation across the subregions of the network, we turned to several additional metrics. First, we examined the extent to which the target output populations from patterns 1 and 2 differed by calculating the percent overlap among the two populations for each network. More formally, we measured the change in population distance via the Hamming distance, which calculates the proportion of positions that differ between two binary vectors:

$$PD\Delta = D_{out} - D_{in} \tag{8}$$

$$D = \frac{1}{N} \sum_{j=1}^{N} X_j \neq Y_j \tag{9}$$

Where $PD\Delta$ is the change in population distance. D_{in} and D_{out} are the Hamming distance between the input and output patterns, respectively. X_i and Y_j are binary vectors representing patterns 1 and 2, and N is the length of the binary vectors. By this metric, identical vectors have a Hamming distance of 0 while two unique vectors have a Hamming distance of 1. If a network performs pattern separation, the Hamming distance of two output populations will be greater than that of the corresponding input populations (Chavlis et al., 2017). If PD $\Delta > 0$, the network performs pattern separation. If $PD\Delta < 0$, the output patterns are more similar than the input patterns. A second feature of pattern separation accounts for rate differences between the output patterns (Chavlis et al., 2017). For this analysis, we focused on the target neurons that were common responders to both patterns and measured the mean Spearman distance between the pattern 1 and pattern 2 responses of common neurons. The Spearman distance (SD) is calculated as one minus the Spearman rank correlation between two vectors:

$$SD = 1 - \frac{(r_s - \bar{r}_s)(r_t - \bar{r}_t)'}{\sqrt{(r_s - \bar{r}_s)(r_s - \bar{r}_s)'}\sqrt{(r_t - \bar{r}_t)(r_t - \bar{r}_t)'}}$$
(10)

$$\overline{r_s} = \frac{1}{N} \sum_{i}^{N} r_{si} = \frac{N+1}{2}$$
(11)

$$\overline{r_t} = \frac{1}{N} \sum_{j}^{N} r_{tj} = \frac{N+1}{2}$$
 (12)

Where r_s and r_t are the rank vectors of x_s and x_t , representing the normalized rate response pattern 1 and pattern 2, respectively. N is the length of the vectors and number of common neurons between patterns 1 and 2.

Statistical analysis

For statistical comparisons between baseline networks and rewired, null models, we used Student's *t*-test. To compare baseline and impaired networks, we applied a paired Student's *t*-test with Bonferroni correction to determine significance for cases of multiple comparison. Statistical testing also included repeated measures ANOVA with Tukey-Kramer *post-hoc* test for comparisons with multiple timepoints.

Results

Modularity in baseline networks

For modularity analysis, we narrowed our focus to areas CA3 and CA1 due to network size and because these two subregions displayed the largest injury effects in our preliminary analysis of functional changes after impairment (Figure 3A). To establish whether the hippocampal networks had detectable community structure (Figure 3B), we compared them to null models generated by rewiring the connections of the original matrix while preserving the input and output degree distributions. We found that the number of modules was significantly lower in the hippocampal model matrices than in the randomized networks, indicating that empirical communities are more integrated than predicted by random models (Baseline hippocampal: 6 \pm 0.5 vs. Randomized: 24.9 \pm 1.5; Student's *t*-test; *p* < 1e-10) (**Figure 3C**). As expected, modularity (Q) was significantly higher in experimental baseline networks than in randomized controls (baseline hippocampal: 0.269 ± 0.002 vs. Randomized: 0.089 ± 0.001 ; Student's *t*-test; p < 1e-10 (Figure 3D). High values of Q mean that the detected communities have higher internal connectivity than predicted by chance. Together, these results confirm that the hippocampal

networks have significant modular structure as compared to null models. Furthermore, we evaluated modularity Q for the last 5 min of simulation time at baseline and found no change in Q over time (One-way ANOVA; F-statistic = 0.08; p > 0.5) (**Figure 3E**). Therefore, we used the final connectivity matrices (from min 30) to compare baseline and impaired networks in subsequent analysis.

Effects of spike-timing-dependent plasticity impairment on community structure

We next compared the community structure of baseline networks with that of STDP impaired networks (Figures 4A,B). Models with STDP impairment ran for an additional 30 min, and the ending connectivity was compared to the pre-injury connectivity using the same modularity algorithm and holding gamma constant. Modularity Q decreased significantly after plasticity impairment (Baseline: 0.26 ± 0.01 vs. STDP Impaired: 0.24 ± 0.02 ; paired Student's *t*-test; p < 0.01) (Figure 4D). However, the number of modules did not differ (Baseline: 5.0 \pm 1.0 vs. STDP Impaired: 5.3 \pm 1.3; Student's *t*-test; *p* > 0.1) (Figure 4E). While the average number of modules per network remained the same, we did identify trends in the sizes of modules after injury. Modules derived from networks with STDP impairment were more likely to fall at the extreme ends of the size range (Figure 4C). In particular, there are more small communities below a size of 250 nodes. On a network level, the size range between the largest and smallest module of each network increased after STDP impairment, reflecting the evolution of these smaller communities (Baseline: 1,129 \pm 333 vs. STDP Impaired: 1,439 \pm 332; Student's *t*-test; p < 0.05) (Figure 4F).

The shifts in module size suggested a broader realignment of neurons among existing communities, and we further hypothesized that the detected community structure might reflect the anatomical designations of the hippocampal circuitry. Therefore, we analyzed the neuron subtype composition of each module for both baseline and impaired networks. Each module was characterized based on the percentage of neurons from CA3 vs. CA1 and the percentage of inhibitory neurons. We found that excitatory neurons from CA3 tended to segregate into their own communities (Figure 5A). The remaining communities contained most of the CA1 excitatory neurons (pyramidal cells) as well as inhibitory neurons from both CA3 and CA1. Accordingly, we identified a significant relationship between the percentage of inhibitory neurons in the module and the percentage of CA1 neurons. As the inhibitory percentage increased, the percentage of CA1 neurons decreased, indicating that these additional inhibitory neurons were anatomically derived from CA3 (Y = 0.40X + 0.005; linear regression; $R^2 = 0.75$; p < 1e-5). After STDP impairment, we found that CA3 excitatory neurons continued to form separate communities; however, the relationship between the percentage of inhibitory neurons and CA1 neurons disappeared (**Figure 5B**). This occurs due to the appearance of many small modules that contain excitatory neurons from both CA1 and CA3. Most likely, some neurons from the CA3 excitatory modules realign with excitatory neurons from CA1 to form these small communities.

Pattern separation in baseline and impaired networks

Learning and memory are crucial hippocampal functions supported by synaptic potentiation. As a mechanism of synaptic weight modification, STDP facilitates use-dependent circuit adaptation. To test whether and how STDP impairment affects higher-level network functions, we implemented a method of unsupervised learning characterized by training-dependent changes in neural activity (Figure 6A). Baseline networks were trained with STDP impairment or under control conditions. During training, two overlapping sets of 200 neurons in the DG were stimulated in addition to receiving baseline noise input. The two stimulus patterns were interleaved and stimulated at 1 Hz. During testing, the same two input patterns were activated in a static network. Networks were tested before and after training to determine the relative change in firing rate on a neuron basis. Not including those neurons stimulated with input patterns, the rest of the principal excitatory neurons in the network were divided into two groups of responders. For each subregion (DG, CA3, and CA1), those that increased their spiking activity the most were termed target neurons, and the remainder were classified as off-target neurons. Target and offtarget neurons were not identified a priori but rather based on their response to the training paradigm.

Although we hypothesized that limiting potentiation would interfere with the encoding phase of memory, we found that both baseline and STDP impaired networks were capable of encoding conditioned responses to input stimulation. The target neurons had significantly higher average normalized firing rate than their off-target counterparts across all three subregions and both conditions (Student's *t*-test; p < 1e-5 for all conditions) (Figures 6B,C). We also computed the signal-to-noise ratio (SNR) as the target activity divided by the off-target activity and found that STDP impaired networks expressed lower SNR in all three subregions of the hippocampus with the most significant change in CA1 (Paired Student's t-test; p < 0.02for all subregions) (Figure 6D). This decrease in SNR appears primarily driven by a decrease in firing among target neurons. Although significant, the magnitude of the difference was modest.

Thus far in our analysis of the learning paradigm, we focused only on the magnitude of the output; however, we



significant change in modularity over time at baseline. *p < 0.01.

also investigated whether the response to each pattern differed. Given the observed decrease in SNR among the responder neurons, we sought to determine whether this decrease affected the ability of the circuit to perform pattern separation by discriminating between the two overlapping input patterns. Successful pattern separation requires that the output patterns be more different than the input patterns (Figure 7A). Accordingly, we evaluated the amount of overlap between the groups of target neurons for each pattern, finding that the mean percentage of overlap was 16 and 13% for the DG and CA1, respectively (Figure 7B). This is well below the 50% overlap of the input patterns, indicating strong pattern separation. Interestingly, the percentage of overlap among target neurons from CA3 was 48% on average (Figure 7B), so this area did not execute pattern separation. This is most likely attributable to the recurrent collaterals in CA3 that putatively make the area uniquely adept at pattern completion, the ability to complete an output response based on partial input information. Due to the limited ability to potentiate synapses, we hypothesized that STDP impairment would limit the ability to encode unique output patterns. However, we found that the percentage of overlap did not decrease in networks that were trained with impairment. In fact, the change in population distance between the input and output populations increased in the DG and CA3 of impaired networks, suggesting that pattern separation was more successful in these subregions (DG: 0.18 ± 0.06 vs. 0.24 ± 0.04 ; paired Student's *t*-test; p < 0.001. CA3: -0.15 ± 0.04 vs. -0.03 ± 0.05 ; p < 0.001. CA1: 0.21 ± 0.03 vs. 0.21 ± 0.02 ; p > 0.1 for CA1) (Figure 7C).

In addition to distinct populations of responsive neurons, rate coding is another attribute of pattern separation (Chavlis et al., 2017). Since most of the neurons in the target populations were unique to one pattern or the other, we were interested in the neurons that activated with both patterns and whether these common neurons responded preferentially to either pattern. We calculated the normalized rate difference between pattern 1 and pattern 2 activity for all common neurons (**Figure 7D**). To compute the distance between the response vectors, we evaluated the mean Spearman distance across networks. We found that the only subregion to show a significant change



significantly after injury (*p < 0.01).

after STDP impairment was CA3, but there were no significant differences in rate coding among common neurons of the DG or CA1 (paired student's *t*-test with Bonferroni correction for multiple comparisons; p < 1e-5 for CA3) (Figure 7E). Therefore, although STDP impairment reduced the total SNR, rate coding was still effective for pattern separation among common responder neurons. While no deficits were observed in population- or rate-based analyses of pattern separation in these circuits, these results do not preclude the possibility that there may be subtle differences in temporal coding based on specific spike timing.

Nodal flexibility in target neurons

Finally, we assessed modularity in trained baseline and STDP impaired networks. Similar to untrained impaired networks, trained circuits with STDP impairment had lower

modularity than untrained baseline networks (Repeated measures ANOVA with Tukey-Kramer post-hoc for multiple comparisons; p < 0.05) (Figure 8A). Trained baseline networks did not significantly differ from either untrained baseline or STDP impaired networks (Figure 8A). After verifying community structure in trained networks, we investigated how community affiliations changed over time. To do so, we applied the concepts of "flexibility" and "promiscuity" (Figure 8B). As it relates to network theory, flexibility describes whether nodes change their community affiliation at different time points. Nodes with high flexibility frequently associate with different modules. Promiscuity is a related yet distinct concept that quantitatively captures whether nodes associate with several unique modules or only a few. A highly flexible node could have low promiscuity if it shifts between only two unique communities. We analyzed flexibility and promiscuity in the aggregate target and off-target populations of CA3 and CA1. Target neurons, which increased their activity the most



FIGURE 5

Module characterization by underlying neuron type reflect hippocampal anatomy. (A) At baseline, one subgroup of modules is comprised primarily of CA3 excitatory neurons (within circle). Predominantly CA1 modules contain most of the inhibitory neurons from both CA3 and CA1. Therefore, there is a significant relationship between the percentage of inhibitory neurons and the percentage of CA1 neurons in these modules (inset) ($R^2 = 0.75$; linear hypothesis test; p < 1e-5). As the percentage of inhibitory neurons increases, the percentage of CA1 neurons decreases (inset). (B) After STDP impairment, there remains a subgroup of modules comprised of CA3 excitatory neurons (within circle). However, a new subgroup of small modules develops. These are made up of excitatory neurons from both CA1 and CA3. The appearance of these small excitatory modules eliminates the relationship between inhibitory tone and the percentage of CA1 neurons (inset) $(R^2 = 0.03;$ linear hypothesis test; p > 0.1). *p < 0.01.

after training, were most likely to fall in the highest or lowest quintiles of the flexibility distribution (**Figure 8C**). These neurons also had low promiscuity, indicating that changes in community assignment included few unique modules (**Figure 8D**). Together, these results suggest that target output neurons have comparatively stable community affiliations since even those that were flexible were associated with lower promiscuity. In contrast, off-target neurons fell relatively evenly into the flexibility and promiscuity quintiles (**Figures 8C,D**). We found no significant differences in these properties after STDP impairment.

Discussion

In these studies, we examined the community structure of a neuronal network model of the hippocampus. At baseline, we found that the CA3-CA1 networks displayed significant modular structure in which excitatory neurons from CA3 (pyramidal cells) reliably segregated into distinct communities. The remaining neurons, including CA1 pyramidal cells and inhibitory neurons from both subregions, formed separate modules. After STDP impairment, modularity decreased significantly, and more small modules appeared. With their small, spurious nature, these modules are purportedly less functionally well-defined. We then trained the networks with an unsupervised learning algorithm to test the critical function of pattern separation across the subregions of the circuit. In the learning process, we identified a critical group of target neurons that showed the largest rate-dependent training effect. STDP impairment during the encoding phase of pattern acquisition reduced the magnitude of the learning effect; however, impaired networks executed pattern separation successfully as analyzed with both population- and rate-based coding. Finally, we found that target neurons had a unique modularity-derived profile characterized by low nodal promiscuity, which indicates that these target neurons were relatively stable and affiliated with few unique network communities. In comparison, offtarget neurons followed more homogenous flexibility and promiscuity distributions.

There are several limitations to the current studies that influence the interpretation of this work. Fundamentally, the hippocampal model is not full-scale and contains a limited number of cell and receptor types. It does not have lamellar structure or complex geometry; however, the synaptic connectivity is faithful to the literature and the most important attribute for the network-based analysis presented here. We use a point neuron model of Izhikevich integrate-and-fire neurons that is more phenomenological than other, more biophysical neuron models. This drawback is balanced by high computational efficiency, which enabled the development of a large network model of the hippocampus, and by extensive use and validation of cell-specific spike timing across different neuron types (Izhikevich, 2003, 2004; Izhikevich and Edelman, 2008; Pena et al., 2018; Muddapu et al., 2019). Similarly, we only modeled some of the ionic currents within each neuron, not including either longer duration NMDA currents (from NR2B containing NMDARs), additional GABAR subtypes, or different types of voltage gated calcium channels. Excluding these currents will alter both the rate and duration of firing frequency, but are less likely to affect the relative change in these patterns which occur when neuronal networks are altered after injury.

Our simulation of STDP impairment as a consequence of mild TBI is also a limitation of these studies. Despite the prevalence of learning and memory deficits after TBI, there is not extensive literature surrounding plasticity impairment. Beyond inhibiting long-term potentiation in hippocampal circuitry, injury reduces CaMKII expression and synaptic protein assemblies, thereby impeding synaptic strengthening (Schwarzbach et al., 2006; Vogel et al., 2017). Given our



or decreases. (C) Networks with STDP impairment exhibit the same paradigm as baseline networks with higher activity in target neurons than in off-target neurons. (D) The signal-to-noise ratio (on-target divided by off-target response) decreases significantly after injury in each region (paired Student's t-test, p < 0.02 with significance determined by Bonferroni correction).

results that STDP impairment did not have a strong negative effect on learning and pattern separation, additional injury mechanisms should be explored in future work. Since damage is known to cause pattern separation deficits in both animals and humans (Brock Kirwan et al., 2012; Paterno et al., 2017; Hanert et al., 2019), our results suggest that some additional mechanism beyond STDP impairment must contribute to those effects. Upcoming modeling studies might also examine the interplay between different plasticity algorithms since a stronger homeostatic mechanism might compensate for larger decreases in STDP-related potentiation, thereby preserving baseline untrained activity levels while exposing larger learning deficits.

Lastly, we implemented an unsupervised learning paradigm, which makes no *a priori* designation between desired and undesired responses. For each of two patterns, we stimulated a subset of 200 neurons in the dentate and identified the most responsive neurons in all three hippocampal subregions based on their normalized firing rate. We also implemented training and STDP impairment at the same time to hold the runtime constant between impaired and control networks. Yet, we could also consider training networks that had already adapted to STDP impairment controlled. It is possible that training mitigated the effects of injury and that networks with ingrained diminished activity are less responsive to training. Although this unsupervised method of network learning cannot address complex temporal coding, it has several advantages. Since it is a computationally efficient post-hoc algorithm without prior topological assumptions, it could be applied with spiking data of this size and density. It also exploits our incorporation of use-dependent plasticity (STDP) as one of the major advances in a model of this size and biological fidelity. Therefore, this method constituted a reasonable biological proxy despite its unsupervised nature. One popular alternative in biologically inspired neural networks is the detection of polychronous neural



FIGURE 7

There is no pattern separation deficit in circuits with STDP impairment. (A) Pattern separation occurs when the output patterns differ more than the input patterns do. In this study, we stimulated two patterns with 50% overlap in the population of input neurons. (B) For each region, the output populations consisted of 200 target neurons for each pattern. The percent overlap in baseline networks was below 20% for the DG and CA1. Similar to baseline networks, STDP impaired networks had low percentage overlap in the DG and CA1 with higher overlap in area CA3. (C) The difference between the Hamming distance of the input population and the output population measures pattern separation where a higher value indicates greater pattern separation. With STDP impairment, the distance between output populations was greater in the DG and CA3 than at baseline (paired Student's t-test, p < 0.02 with significance determined by Bonferroni correction). (D) The rate difference between one neurons shows that common neurons responded preferentially to one pattern or the other. Common target neurons from the DG in one representative network are shown. P1 = pattern 1; P2 = pattern 2. (E) The distance between the rate response to pattern 1 vs. pattern 2 was computed as the Spearman distance. The rate distance for CA3 outputs was significantly different between baseline and STDP impaired networks (paired Student's t-test, p < 10.2).



groups, which is better adapted to handling many neural groups and memory traces and evaluating the maximal amount of information that might be stored in a given circuit. While the original algorithm requires computationally expensive, bruteforce computations, some groups are developing more efficient alternatives inspired by the field of machine learning (Guise et al., 2014; Chrol-Cannon et al., 2017). These approaches might offer an opportunity to extend these results with a quantification of the information storage capacity of this hippocampal circuit.

Modularity is a useful framework for assessing the architectural organization of a network. Large networks often consist of several smaller subnetworks that are more densely connected internally than they are externally (Sporns and Betzel, 2016). This partitioned organization is posited to support faster, more efficient processing by facilitating functional compartmentalization (Sporns and Betzel, 2016; Khambhati et al., 2018). By reducing the energy requirements for networkwide modifications, a more modular structure is also a more adaptable one (Bassett et al., 2011; Arnemann et al., 2015). In the present study, we identify post-injury modularity reduction, which may constitute an adaptation that increases integration between communities to support overall activity levels. This result further suggests that potentiation supports baseline segregation in the hippocampal circuit. Although there is also evidence of the opposite (Han et al., 2014, 2020),

previous findings that TBI reduces modularity in functional brain networks correlate with persistent post-concussive syndrome (Messé et al., 2013; Arnemann et al., 2015). In addition, more modular structure was predictive of better training outcomes after injury (Arnemann et al., 2015); this may occur because lower energy costs are associated with adaptation in more highly segregated networks. The variable response (increase vs. decrease) may relate to individual heterogeneity, other measures of network-wide integration, or whether the network is still in a state of active adaptation. Aside from analyzing the microcircuit scale, differences between our results and others may be attributable to our focus on structural, instead of functional, connectivity. Since functional connectivity is dynamic and malleable while structural connectivity is more stable as a reflection of the underlying neural anatomy, it is possible that the training effect is larger for functional networks. Ultimately, the demonstrated effects in structurally well-defined microcircuits corroborate the idea that modularity may be a useful (bio)marker of intervention-dependent network plasticity (Gallen and D'Esposito, 2019).

With a modest amount decrement in STDP-related potentiation, networks could still learn and execute pattern separation. In another recent study from our group, we tested the circuit-level consequences of NMDA receptor damage, which increases network activity, in a generic circuit with a similar learning paradigm (Gabrieli et al., 2020a). Since injuryinduced, elevated activity obscured the training effect, we found the most detrimental outcome of NMDA receptor dysfunction occurred during recall of previously trained patterns but also tested injury during different phases of memory (Gabrieli et al., 2020a). Here, we exclusively tested STDP impairment during the encoding stage. In our preliminary work analyzing impaired networks without exogenous stimulation, we found significant declines in firing rate and signal power. Based on those effects, we expect the maintenance phase would also challenge STDP impaired networks, which would likely lose the entrained response more quickly as overall activity decreases without exogenous stimulation. This idea is supported by behavior studies that find injured animals perform the task successfully if tested quickly after training but not if the time between testing and training is longer (Paterno et al., 2018). If we integrated both STDP impairment and NMDA receptor damage simultaneously, we expect that STDP impairment might enhance pattern recall because the two mechanisms have opposing influence on network activity. Alternatively, as trauma-induced changes to NMDAR physiology will disappear when receptors are replaced hours after injury (Giza et al., 2006; Reger et al., 2012; Estrada-Rojo et al., 2018) and plasticity impairments may persist for days after injury, one might expect an acute early impairment in the retrograde memory (Smith et al., 1991; Gorman et al., 1993; Whiting and Hamm, 2008) with a longer lasting impairment in memory acquisition (An et al., 2016; Luo et al., 2017; Paterno et al., 2018).

As a measure of encoding, pattern separation conveys the capacity to distinguish similar events and contexts; therefore, this function underpins general learning abilities. TBI causes behavioral deficits in spatial memory and spatial object recognition in animal models of injury (Paterno et al., 2017, 2018; Folweiler et al., 2018). Moreover, using a standard T-maze behavior paradigm, injured mice showed impaired working memory up to 7 days post-injury, suggesting that TBI interferes with the process of memory formation (Smith et al., 2015). These trained behaviors depend on discrimination between similar experiences. Recent findings also demonstrate that injury impairs pattern separation in humans (Brock Kirwan et al., 2012; Hanert et al., 2019). The dentate is traditionally the primary focus of studies on hippocampal pattern separation because its intrinsic properties of high inhibition and parallel circuity intuitively support this filter function; however, there is growing evidence that other subregions (CA3 and CA1) also facilitate pattern separation. In fact, temporary CA1 lesions impair pattern separation in humans (Hanert et al., 2019). Since CA1 relays information processed by the hippocampus to neocortical brain regions (Witter and Amaral, 2004), the area clearly plays an important role in the wider circuitry, making it an intuitively important subregion. For these reasons, it is interesting that in our work changes in the DG and CA3 appear to compensate for one another because the population- and rate-based output distances measured from CA1 do not differ. These results suggest that the output patterns transduced by CA1 are essentially the same and that the network adapts to maintain that final output. While one might predict a larger effect of STDP impairment on pattern separation, these subtle differences are an intuitive extension of our collective results. With our preliminary functional analysis in networks without learning, we found that the DG was remarkably robust after STDP impairment. Specifically, the average firing rate and signal power did not decrease significantly. Given its intrinsically low rates of activity, the DG is more resilient to minor changes in STDP. Others have found that deficits in pattern separation are associated with hyperexcitability and elevated activity in the DG (Myers and Scharfman, 2009; Jinde et al., 2012; Chavlis et al., 2017), which increases activation and thereby reduces the capacity of the filter function. In general, STDP impairment reduces synaptic weights in the network, making it more difficult to activate. Although its impact on learning is more indirect, NMDA receptor damage or inhibitory cell degeneration might have outsized influence on pattern separation because these mechanisms would increase spurious noise in the output patterns. Finally, our analysis in this work focused on population and rate coding; however, we cannot exclude the importance of temporal coding because it is possible that the spike timing changes while the activity rate remains stable. Indeed, previous results from our group indicate that networks adapt to preserve firing rate first as other measures of spike timing exhibit longer lasting changes after neurodegeneration (Gabrieli et al., 2020b).

One natural extension of this work is prospective training or other interventions designed to facilitate active recovery in damaged networks. For instance, a stimulation protocol that could restore activity in a damaged network would be of interest for rehabilitation (Pevzner et al., 2016; Paterno et al., 2017), and certain types of stimulation (frequencies, magnitudes, etc.) might bet associated with better training outcomes. There is a clear need to investigate stimulation in conjunction with injury and the role that it may play in network recovery. It is often assumed that concussed patients should limit exposure to any form of stimulation because it mitigates their symptoms; however, targeted stimulation may instead help resolve chronic deficits (Pevzner et al., 2016). Relatedly, the functional connectivity characteristics of our hippocampal network should be examined more completely, as we may discover a structurally modified network achieves nearly the same functional organization that appeared before injury. This analysis would enable us to address how closely functional networks reflect underlying structural connectivity at the microcircuit scale. At the macroscale, a link between axonal tractography and a resting state functional network is established (Greicius et al., 2009; Honey et al., 2009), but the relationship between structural and functional connectivity is not well-understood in microcircuits. Further, characterizing functional networks from these simulations would offer an opportunity to link this work with experimental results measured via microelectrode arrays and make structurally based insights about those empirical functional data (Kang et al., 2015).

With this work, we investigate the modular network structure of a computational model of the hippocampus, a region of the brain that has well-characterized anatomy and electrophysiology, and we examine the functional implications of plasticity impairment on network-defined pattern separation. These studies contribute to a growing body of work regarding the circuit-level effects of cellular damage in neuronal networks (Volman et al., 2011; Gabrieli et al., 2020a,b; Schumm et al., 2020). Studying a posited substrate of physiological learning with this biologically inspired computational model of the hippocampus, which is known for its role in learning and memory, guides new insights into both temporary and more permanent impairments that could occur from cellular-based changes after traumatic injury. In addition, combining this cellular-level mechanistic insight with new tools in data science (e.g., deep learning and machine learning) provides an opportunity to create biologically inspired autonomous learning models that could aid the recovery and repair of damaged circuits. By understanding network-based learning in

this hippocampal circuit, we will not only advance practical analytical tools, but we may also develop targeted interventions to improve outcomes for patients with diseases of brain-network organization.

Data availability statement

Publicly available datasets were analyzed in this study. This data can be found here: https://www.seas.upenn.edu/ ~{}molneuro/research.html.

Author contributions

SS and DM conceived and designed the studies and analysis, and wrote the manuscript. SS performed simulations and conducted analysis. SS and DG contributed model and analysis tools. DG contributed to the manuscript discussion and revision. All authors contributed to the article and approved the submitted version.

Funding

Funding provided by grants by the Paul G. Allen Foundation and the NIH (RO1 NS088176).

Acknowledgments

The following figures or figure panels were created with BioRender.com: Figures 1A,B, 2, 6A, 7A.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

References

Aika, Y., Ren, J. Q., Kosaka, K., and Kosaka, T. (1994). Quantitative analysis of GABA-like-immunoreactive and parvalbumin-containing neurons in the CA1 region of the rat hippocampus using a stereological method, the disector. *Exp. Brain Res.* 99, 267–276. doi: 10.1007/BF00239593

Albensi, B. C., Sullivan, P. G., Thompson, M. B., Scheff, S. W., and Mattson, M. P. (2000). Cyclosporin ameliorates traumatic brain-injury-induced alterations of hippocampal synaptic plasticity. *Exp. Neurol.* 162, 385–389. doi: 10.1006/exnr. 1999.7338

An, C., Jiang, X., Pu, H., Hong, D., Zhang, W., Hu, X., et al. (2016). Severitydependent long-term spatial learning-memory impairment in a mouse model of traumatic brain injury. *Transl. Stroke Res.* 7, 512–520. doi: 10.1007/s12975-016-0483-5

Arnemann, K. L., Chen, A. J. W., Novakovic-Agopian, T., Gratton, C., Nomura, E. M., and D'Esposito, M. (2015). Functional brain network modularity predicts response to cognitive training after brain injury. *Neurology* 84, 1568–1574.

Aungst, S. L., Kabadi, S. V., Thompson, S. M., Stoica, B. A., and Faden, A. I. (2014). Repeated mild traumatic brain injury causes chronic neuroinflammation, changes in hippocampal synaptic plasticity, and associated cognitive deficits. *J. Cereb. Blood Flow. Metab.* 34, 1223–1232. doi: 10.1038/jcbfm.2014.75

Bassett, D. S., Wymbs, N. F., Porter, M. A., Mucha, P. J., Carlson, J. M., and Grafton, S. T. (2011). Dynamic reconfiguration of human brain networks during learning. *Proc. Natl. Acad. Sci. U.S.A.* 108, 7641–7646.

Bezaire, M. J., and Soltesz, I. (2013). Quantitative assessment of CA1 local circuits: Knowledge base for interneuron-pyramidal cell connectivity. *Hippocampus* 23, 751–785. doi: 10.1002/hipo.22141

Bezaire, M. J., Raikov, I., Burk, K., Vyas, D., and Soltesz, I. (2016). Interneuronal mechanisms of hippocampal theta oscillations in a full-scale model of the rodent CA1 circuit. *Elife* 5:e18566. doi: 10.7554/eLife.18566

Blondel, V. D., Guillaume, J. L., Lambiotte, R., and Lefebvre, E. (2008). Fast unfolding of communities in large networks. *J. Stat. Mech. Theory Exp.* 2008:10008. doi: 10.1103/PhysRevE.83.036103

Brock Kirwan, C., Hartshorn, A., Stark, S. M., Goodrich-Hunsaker, N. J., Hopkins, R. O., and Stark, C. E. L. (2012). Pattern separation deficits following damage to the hippocampus. *Neuropsychologia* 50, 2408–2414. doi: 10.1016/j. neuropsychologia.2012.06.011

Cancelliere, C., Coronado, V. G., Taylor, C. A., and Xu, L. (2017). Epidemiology of isolated versus nonisolated mild traumatic brain injury treated in emergency departments in the United States, 2006-2012: Sociodemographic characteristics. *J. Head Trauma Rehabil.* 32, E37–E46. doi: 10.1097/HTR.00000000000260

Caporale, N., and Dan, Y. (2008). Spike timing-dependent plasticity: A Hebbian learning rule. *Annu. Rev. Neurosci.* 31, 25–46.

Chavlis, S., Petrantonakis, P. C., and Poirazi, P. (2017). Dendrites of dentate gyrus granule cells contribute to pattern separation by controlling sparsity. *Hippocampus* 27, 89–110.

Chrol-Cannon, J., Jin, Y., and Grüning, A. (2017). An efficient method for online detection of polychronous patterns in spiking neural networks. *Neurocomputing* 267, 644–650.

Cohen, A. S., Pfister, B. J., Schwarzbach, E., Sean Grady, M., Goforth, P. B., and Satin, L. S. (2007). Injury-induced alterations in CNS electrophysiology. *Prog. Brain Res.* 161, 143–169. doi: 10.1016/S0079-6123(06)61010-8

Dawish, H., Mahmood, A., Schallert, T., Chopp, M., and Therrien, B. (2012). Mild traumatic brain injury (MTBI) leads to spatial learning deficits. *Brain Inj.* 26, 151–165.

Draguhn, A., and Buzsáki, G. (2004). Neuronal oscillations in cortical networks. Science 304, 1926–1930.

Effenberger, F., Jost, J., and Levina, A. (2015). Self-organization in balanced state networks by STDP and homeostatic plasticity. *PLoS Comput. Biol.* 11:e1004420. doi: 10.1371/journal.pcbi.1004420

Estrada-Rojo, F., Morales-Gomez, J., Coballase-Urrutia, E., Martinez-Vargas, M., and Navarro, L. (2018). Diurnal variation of NMDA receptor expression in the rat cerebral cortex is associated with traumatic brain injury damage. *BMC Res. Notes* 11:150. doi: 10.1186/s13104-018-3258-0

Feldman, D. E. (2012). The spike-timing dependence of plasticity. *Neuron* 75, 556–571.

Folweiler, K. A., Samuel, S., Metheny, H. E., and Cohen, A. S. (2018). Diminished dentate gyrus filtering of cortical input leads to enhanced area CA3 excitability after mild traumatic brain injury. *J. Neurotrauma* 35, 1304–1317. doi: 10.1089/ neu.2017.5350

Gabrieli, D., Schumm, S. N., Vigilante, N. F., and Meaney, D. F. (2020a). NMDA receptor alterations after mild traumatic brain injury induce deficits in memory acquisition and recall. *Neural. Comput.* 33, 67–95. doi: 10.1162/neco_a_01343

Gabrieli, D., Schumm, S. N., Vigilante, N. F., Parvesse, B., and Meaney, D. F. (2020b). Neurodegeneration exposes firing rate dependent effects on oscillation dynamics in computational neural networks. *PLoS One* 15:e0234749. doi: 10.1371/journal.pone.0234749.g001

Gallen, C. L., and D'Esposito, M. (2019). Brain modularity: A biomarker of intervention-related plasticity. *Trends Cogn. Sci.* 23, 293–304. doi: 10.1016/j.tics. 2019.01.014

Gaudette, É, Seabury, S. A., Temkin, N., Barber, J., Digiorgio, A. M., Markowitz, A. J., et al. (2022). Employment and Economic Outcomes of Participants With Mild Traumatic Brain Injury in the TRACK-TBI Study. *JAMA Netw. Open* 5:e2219444. doi: 10.1001/jamanetworkopen.2022.19444

Genon, S., Bernhardt, B. C., La Joie, R., Amunts, K., and Eickhoff, S. B. (2021). The many dimensions of human hippocampal organization and (dys)function. *Trends Neurosci.* 44, 977–989. doi: 10.1016/j.tins.2021.10.003

Giza, C. C., Prins, M. L., and Hovda, D. A. (2017). It's not all fun and games: Sports. Concussions, and Neuroscience. *Neuron* 94, 1051–1055.

Giza, C. C., Santa Maria, N. S., and Hovda, D. A. (2006). N-methyl-Daspartate receptor subunit changes after traumatic injury to the developing brain. *J. Neurotrauma* 23, 950–961. doi: 10.1089/neu.2006.23.950

Gorman, L. K., Shook, B. L., and Becker, D. P. (1993). Traumatic brain injury produces impairments in long-term and recent memory. *Brain Res.* 614, 29–36.

Greicius, M. D., Supekar, K., Menon, V., and Dougherty, R. F. (2009). Restingstate functional connectivity reflects structural connectivity in the default mode network. *Cereb. Cortex* 19, 72–78.

Guise, M., Knott, A., and Benuskova, L. (2014). A bayesian model of polychronicity. *Neural. Comput.* 26, 2052–2073. doi: 10.1162/NECO_a_00620

Guise, M., Knott, A., and Benuskova, L. (2015). Enhanced polychronization in a spiking network with metaplasticity. *Front. Comput. Neurosci.* 9:9. doi: 10.3389/fncom.2015.00009

Han, K., Chapman, S. B., and Krawczyk, D. C. (2020). Cognitive training reorganizes network modularity in traumatic brain injury. *Neurorehabil. Neural. Repair* 34, 26–38.

Han, K., Mac Donald, C. L., Johnson, A. M., Barnes, Y., Wierzechowski, L., Zonies, D., et al. (2014). Disrupted modular organization of resting-state cortical functional connectivity in U.S. military personnel following concussive "mild" blast-related traumatic brain injury. *Neuroimage* 84, 76–96. doi: 10.1016/j. neuroimage.2013.08.017

Hanert, A., Pedersen, A., and Bartsch, T. (2019). Transient hippocampal CA1 lesions in humans impair pattern separation performance. *Hippocampus* 29, 736–747.

Honey, C. J., Sporns, O., Cammoun, L., Gigandet, X., Thiran, J. P., Meuli, R., et al. (2009). Predicting human resting-state functional connectivity from structural connectivity. *Proc. Natl. Acad. Sci. U.S.A.* 106, 2035–2040.

Izhikevich, E. M. (2003). Simple model of spiking neurons. *IEEE Trans. Neural. Netw.* 14, 1569–1572.

Izhikevich, E. M. (2004). Which model to use for cortical spiking neurons? *IEEE Trans. Neural. Netw.* 15, 1063–1070.

Izhikevich, E. M. (2006). Polychronization: Computation with spikes. Neural. Comput. 18, 245-282.

Izhikevich, E. M., and Edelman, G. M. (2008). Large-scale model of mammalian thalamocortical systems. *Proc. Natl. Acad. Sci. U.S.A.* 105, 3593–3598.

Jeffery, K. J. (2018). The hippocampus: From memory, to map, to memory map. *Trends Neurosci.* 41, 64–66.

Jinde, S., Zsiros, V., Jiang, Z., Nakao, K., Pickel, J., Kohno, K., et al. (2012). Hilar mossy cell degeneration causes transient dentate granule cell hyperexcitability and impaired pattern separation. *Neuron* 76, 1189–1200. doi: 10.1016/j.neuron.2012. 10.036

Josselyn, S. A., Köhler, S., and Frankland, P. W. (2015). Finding the engram. Nat. Rev. Neurosci. 16, 521–534.

Kang, W. H., Cao, W., Graudejus, O., Patel, T. P., Wagner, S., Meaney, D. F., et al. (2015). Alterations in hippocampal network activity after in vitro traumatic brain injury. *J. Neurotrauma* 32, 1011–1019.

Khambhati, A. N., Mattar, M. G., Wymbs, N. F., Grafton, S. T., and Bassett, D. S. (2018). Beyond modularity: Fine-scale mechanisms and rules for brain network reconfiguration. Neuroimage 166, 385-399. doi: 10.1016/j.neuroimage.2017.11. 015

Kondziella, D., Amiri, M., Othman, M. H., Beghi, E., Bodien, Y. G., Citerio, G., et al. (2022). Incidence and prevalence of coma in the UK and the USA. *Brain Commun.* 4:fcac188.

Kotapka, M. J., Graham, D. I., Adams, J. H., and Gennarelli, T. A. (1994). Hippocampal pathology in fatal human head injury without high intracranial pressure. *J. Neurotrauma* 11, 317–324.

Lopez, D. A., Christensen, Z. P., Foxe, J. J., Ziemer, L. R., Nicklas, P. R., and Freedman, E. G. (2022). Association between mild traumatic brain injury, brain structure, and mental health outcomes in the adolescent brain cognitive development study. *Neuroimage* 263:119626.

Lu, J. T., Li, C. Y., Zhao, J. P., Poo, M. M., and Zhang, X. H. (2007). Spike-timingdependent plasticity of neocortical excitatory synapses on inhibitory interneurons depends on target cell type. *J. Neurosci.* 27, 9711–9720.

Luo, Y., Zou, H., Wu, Y., Cai, F., Zhang, S., and Song, W. (2017). Mild traumatic brain injury induces memory deficits with alteration of gene expression profile. *Sci. Rep.* 7:10846.

Mariajoseph, F. P., Chen, Z., Sekhar, P., Rewell, S. S., O'Brien, T. J., Antonic-Baker, A., et al. (2022). Incidence and risk factors of posttraumatic epilepsy following pediatric traumatic brain injury: A systematic review and meta-analysis. *Epilepsia*. 63, 2802–2812. doi: 10.1111/epi.17398

McAllister, T. W. (2011). Neurobiological consequences of traumatic brain injury. *Dialogues Clin. Neurosci.* 13, 287–300.

Mcallister, T. W., Sparling, M. B., Flashman, L. A., Guerin, S. J., Mamourian, A. C., and Saykin, A. J. (2001). Differential working memory load effects after mild traumatic brain injury. *Neuroimage* 14, 1004–1012. doi: 10.1006/nimg.2001.0899

Messé, A., Caplain, S., Pélégrini-Issac, M., Blancho, S., Lévy, R., Aghakhani, N., et al. (2013). Specific and evolving resting-state network alterations in postconcussion syndrome following mild traumatic brain injury. *PLoS One* 8:65470. doi: 10.1371/journal.pone.0065470

Muddapu, V. R., Mandali, A., Chakravarthy, V. S., and Ramaswamy, S. (2019). A computational model of loss of dopaminergic cells in parkinson's disease due to glutamate-induced excitotoxicity. *Front. Neural. Circuits* 13:11. doi: 10.3389/fncir. 2019.00011

Myers, C. E., and Scharfman, H. E. (2009). A Role for hilar cells in pattern separation in the dentate gyrus: A computational approach. *Hippocampus* 19, 321–337. doi: 10.1002/hipo.20516

Nicholl, J., and Curt LaFrance, W. (2009). Neuropsychiatric sequelae of traumatic brain injury. Semin. Neurol. 29, 247–255.

Paterno, R., Folweiler, K. A., and Cohen, A. S. (2017). Pathophysiology and treatment of memory dysfunction after traumatic brain injury. *Curr. Neurol. Neurosci. Rep.* 17:52.

Paterno, R., Metheny, H., and Cohen, A. S. (2018). Memory deficit in an object location task after mild TBI is associated with impaired early object exploration and both are restored by branched chain amino acid dietary therapy. *J. Neurotrauma* 35, 2117–2124. doi: 10.1089/neu.2017.5170

Pena, R. F. O., Zaks, M. A., and Roque, A. C. (2018). Dynamics of spontaneous activity in random networks with multiple neuron subtypes and synaptic noise: Spontaneous activity in networks with synaptic noise. *J. Comput. Neurosci.* 45, 1–28. doi: 10.1007/s10827-018-0688-6

Penn, Y., Segal, M., and Moses, E. (2016). Network synchronization in hippocampal neurons. Proc. Natl. Acad. Sci. U.S.A. 113, 3341-3346.

Pevzner, A., Izadi, A., Lee, D. J., Shahlaie, K., and Gurkoff, G. G. (2016). Making waves in the brain: What are oscillations, and why modulating them makes sense for brain injury. *Front. Syst. Neurosci.* 10:30. doi: 10.3389/fnsys.2016.00030

Prince, C., and Bruhns, M. E. (2017). Evaluation and treatment of mild traumatic brain injury: The role of neuropsychology. *Brain Sci.* 7:105.

Raghupathi, R. (2004). Cell death mechanisms following traumatic brain injury. *Brain Pathol.* 14, 215–222.

Reger, M. L., Poulos, A. M., Buen, F., Giza, C. C., Hovda, D. A., and Fanselow, M. S. (2012). Concussive brain injury enhances fear learning and excitatory processes in the amygdala. *Biol. Psychiatry* 71, 335–343. doi: 10.1016/j.biopsych. 2011.11.007

Rubinov, M., and Sporns, O. (2010). Complex network measures of brain connectivity: Uses and interpretations. *Neuroimage* 52, 1059–1069.

Ruff, R. (2005). Two decades of advances in understanding of mild traumatic brain injury. J. Head Trauma Rehabil. 20, 5–18.

Sanders, H., Rennó-Costa, C., Idiart, M., and Lisman, J. (2015). Grid cells and place cells: An integrated view of their navigational and memory function. *Trends Neurosci.* 38, 763–775. doi: 10.1016/j.tins.2015.10.004

Sanders, M. J., Sick, T. J., Perez-Pinzon, M. A., Dietrich, W. D., and Green, E. J. (2000). Chronic failure in the maintenance of long-term potentiation following fluid percussion injury in the rat. *Brain Res.* 861, 69–76. doi: 10.1016/s0006-8993(00)01986-7

Schumm, S. N., Gabrieli, D., and Meaney, D. F. (2020). Neuronal degeneration impairs rhythms between connected microcircuits. *Front. Comput. Neurosci.* 14:18. doi: 10.3389/fncom.2020.00018

Schumm, S. N., Gabrieli, D., and Meaney, D. F. (2022). Plasticity impairment exposes CA3 vulnerability in a hippocampal network model of mild traumatic brain injury. *Hippocampus* 32, 231–250. doi: 10.1002/hipo. 23402

Schwarzbach, E., Bonislawski, D. P., Xiong, G., and Cohen, A. S. (2006). Mechanisms underlying the inability to induce area CA1 LTP in the mouse after traumatic brain injury. *Hippocampus* 16, 541–550. doi: 10.1002/hipo. 20183

Slotnick, S. D. (2022). The hippocampus and long-term memory. Cogn. Neurosci. 13, 113–114. doi: 10.1080/17588928.2022.2128736

Smith, C. J., Xiong, G., Elkind, J. A., Putnam, B., and Cohen, A. S. (2015). Brain injury impairs working memory and prefrontal circuit function. *Front. Neurol.* 6:240. doi: 10.3389/fneur.2015.00240

Smith, D. H., Okiyama, K., Thomas, M. J., Claussen, B., and McIntosh, T. K. (1991). Evaluation of memory dysfunction following experimental brain injury using the morris water maze. *J. Neurotrauma* 8, 259–269.

Song, S., Miller, K. D., and Abbott, L. F. (2000). Competitive Hebbian learning through spike-timing-dependent synaptic plasticity. *Nat. Neurosci.* 3, 919–926.

Sporns, O., and Betzel, R. F. (2016). Modular brain networks. Annu. Rev. Psychol. 67, 613-640.

Taylor, C. A., Bell, J. M., Breiding, M. J., and Xu, L. (2017). Traumatic brain injury-related emergency department visits, hospitalizations, and deaths — United States, 2007 and 2013. *MMWR Surveill. Summ.* 66, 1–16. doi: 10.15585/mmwr.ss6609a1

Tetzlaff, C., Kolodziejski, C., Timme, M., and Wörgötter, F. (2011). Synaptic scaling in combination with many generic plasticity mechanisms stabilizes circuit connectivity. *Front. Comput. Neurosci.* 5:47. doi: 10.3389/fncom.2011.00047

Tulving, E. (1985). How many memory systems are there? Am. Psychol. 40, 385-398.

Turrigiano, G. G., and Nelson, S. B. (2004). Homeostatic plasticity in the developing nervous system. *Nat. Rev. Neurosci.* 5, 97–107.

Turrigiano, G. G., Leslie, K. R., Desai, N. S., Rutherford, L. C., and Nelson, S. B. (1998). Activity-dependent scaling of quantal amplitude in neocortical neurons. *Nature* 391, 892–896.

Vogel, E. W., Rwema, S. H., Meaney, D. F., Bass, C. R. Q., and Morrison, B. (2017). Primary blast injury depressed hippocampal long-term potentiation through disruption of synaptic proteins. *J. Neurotrauma* 34, 1063–1073. doi: 10. 1089/neu.2016.4578

Volman, V., Bazhenov, M., and Sejnowski, T. J. (2011). Pattern of trauma determines the threshold for epileptic activity in a model of cortical deafferentation. *Proc. Natl. Acad. Sci. U.S.A.* 108, 15402–15407. doi: 10.1073/pnas. 1112066108

White, E. R., Pinar, C., Bostrom, C. A., Meconi, A., and Christie, B. R. (2017). Mild traumatic brain injury produces long-lasting deficits in synaptic plasticity in the female juvenile hippocampus. *J. Neurotrauma* 34, 1111–1123. doi: 10.1089/ neu.2016.4638

Whiting, M. D., and Hamm, R. J. (2008). Mechanisms of anterograde and retrograde memory impairment following experimental traumatic brain injury. *Brain Res.* 1213, 69–77. doi: 10.1016/j.brainres.2008.01.107

Whittington, M. A., and Traub, R. D. (2003). Interneuron Diversity series: Inhibitory interneurons and network oscillations in vitro. *Trends Neurosci.* 26, 676–682.

Wilson, N. M., Titus, D. J., Oliva, A. A., Furones, C., and Atkins, C. M. (2016). Traumatic brain injury upregulates phosphodiesterase expression in the hippocampus. *Front. Syst. Neurosci.* 10:5. doi: 10.3389/fnsys.2016.00005

Witter, M. P., and Amaral, D. G. (2004). "Hippocampal formation," in *The rat nervous system*, (Amsterdam: Elsevier Inc), 635–704.

Woodson, W., Nitecka, L., and Ben-Ari, Y. (1989). Organization of the GABAergic system in the rat hippocampal formation: A quantitative immunocytochemical study. *J. Comp. Neurol.* 280, 254–271. doi: 10.1002/cne. 902800207

Zenke, F., Gerstner, W., and Ganguli, S. (2017). The temporal paradox of Hebbian learning and homeostatic plasticity. *Curr. Opin Neurobiol.* 43, 166–176. doi: 10.1016/j.conb.2017.03.015